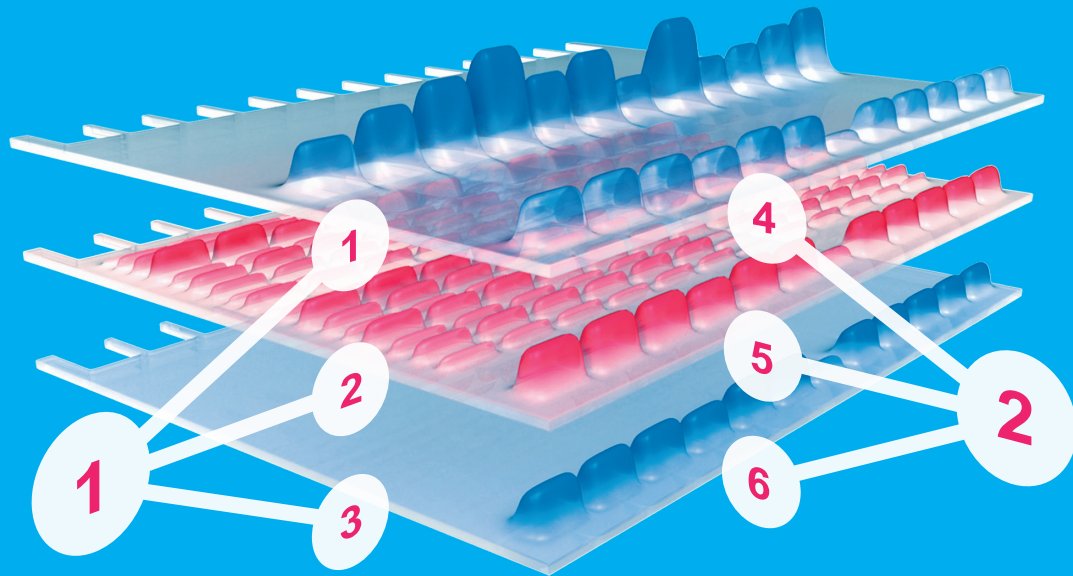


# SMART PROTEIN LAYERS

Quantitative and  
standardized protein gel  
and Western blot analysis



## The challenge

There is an increasing demand for quantitative, sensitive and reliable 1D gel and Western blot (WB) analyses. However, standardization, normalization and precise quantification of different protein samples separated by 1D gel electrophoresis is difficult due for various reasons. Visualization of proteins using stains like *Coomassie Blue* for gels or *Ponceau Red* for blots suffers from low sensitivity, limited dynamic range and poor reproducibility.

Obtaining quantitative data from WB requires an appropriate method of normalization to assure that the reported fold changes of the target protein are not an artifact of reference signal and experimental errors\*.

A typical approach is to use signal normalization of target based on housekeeping proteins (HKPs) like  $\beta$ -actin,  $\beta$ -tubulin or GAPDH. However, as it becomes clear that HKPs are differing in their abundance under various conditions (e.g. stress, treatments), this approach may no longer be suitable.

The use of total protein transferred to the blot is a more accurate source of data normalization\*\*. As proteins are washed away from the membrane during WB procedure, target and reference proteins should be detected at the same moment to achieve accurate quantification.

\* Taylor and Posch. 2014. The design of a quantitative Western blot experiment. BioMed Research International. Article ID 361590

\*\* Colella et al. 2012. Comparison of stain-free gels with traditional immunoblot loading control. Anal. Biochem. 430 (2): 108-110

\*\* Guertler et al. 2013. Stain-free technology as normalization tool in western blot analysis. Anal. Biochem. 433 (2): 105-111

## The solution

Smart Protein Layers (SPL) is the new technology for stain-free, quantitative and standardized analysis of protein gels and Western blots. For the first time the amount of target protein and the amount of total protein of the sample (as a reliable reference) present at this moment can be detected. In addition, SPL allows to precisely compare and quantify data derived from different experiments.

SPL offers:

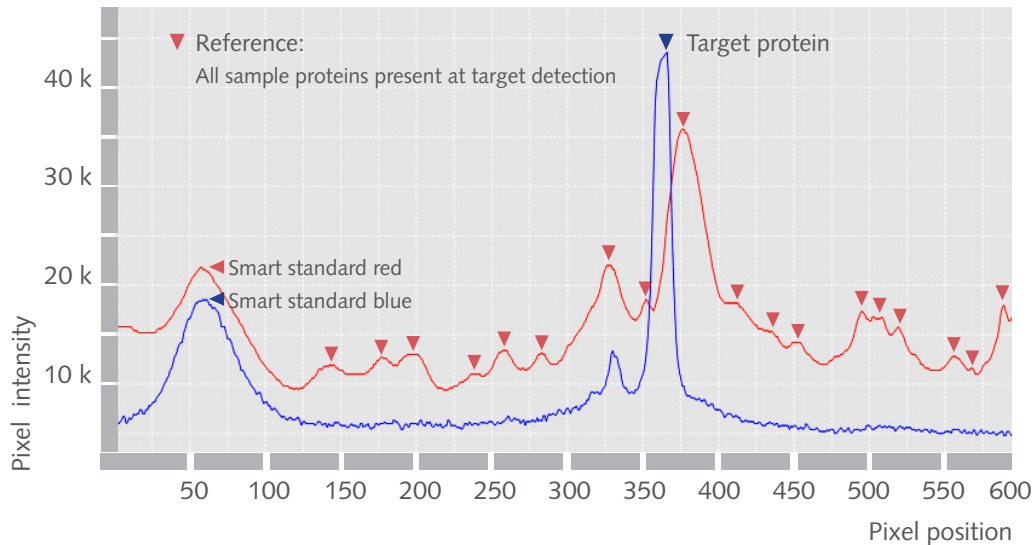
- Stain-free, very sensitive protein visualization on gels and blots
- Protein content equalizer (loading by volume, inaccurate protein determination, loss of sample)
- Monitoring of every step during the Western blot analysis
- Accurate quantification of target protein expression
- Precise comparison of different experiments

The SPL technology is based on fluorescent label for total protein detection combined with a sample-dependent bi-fluorescent standard. This combination allows for rapid and highly sensitive protein detection with a new quality of standardization and accurate quantification of expression levels within one experiment and between different experiments.

# Smart Protein Layers

The first standard-based technique for quantitative Western blots.

## Band profile of a SPL total protein lane at the moment of target detection

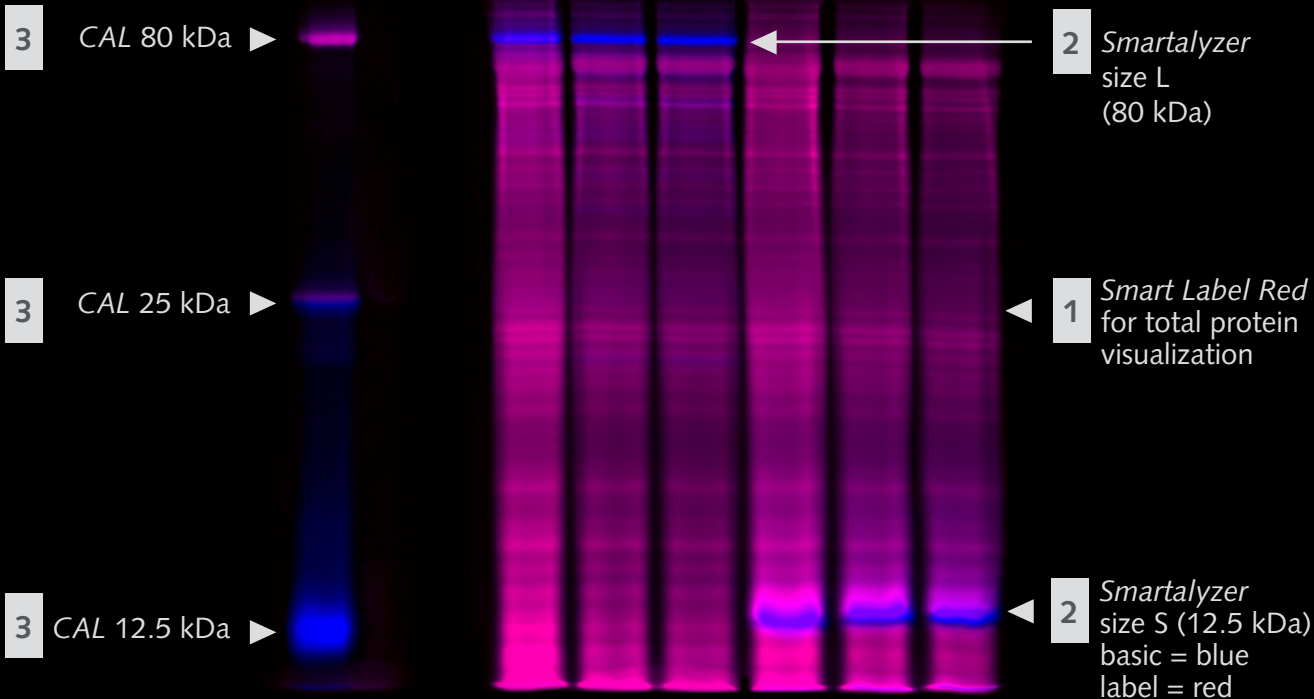


## Smart Protein Layers

### Quantitative and standardized protein gel and Western blot analysis

Smart Protein Layers (SPL) is based on three components:

- 1 The fluorescent *Smart Label* visualizes total protein in the gel and on the blot with high sensitivity (detection limit less than 1 ng, dynamic range  $10^4$  -  $10^5$ ) within seconds. *Smart Labels* are chemically bound to the protein prior protein separation (3 min hands-on).
- 2 The bi-fluorescent *Smartalyzer* (SMA) is a multi-functional standard added to every sample prior to separation. For precise normalization, standardization and quantification of total protein. SMA is available in size S (12.5 kDa) and size L (80 kDa).
- 3 The bi-fluorescent *Calibrator* (CAL) is the standard for comparison protein expression between different gels or blots. CAL also works as bi-fluorescent molecular weight marker (80 kDa; 25 kDa; 12.5 kDa).

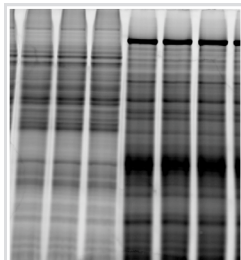


## The Smart Label

### Stain-free and sensitive protein visualization of gels and blots

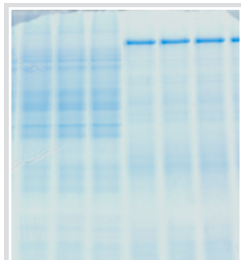
The fluorescent *Smart Label* visualizes total protein in the gel and on the blot with high sensitivity (detection limit less than 1 ng, dynamic range  $10^4 - 10^5$ ) within seconds. *Smart Labels* are chemically bound to the protein prior protein separation (3 min hands-on).

Smart Label



20 µg protein / lane

Coomassie stain



20 µg protein / lane

|                      | Coomassie stain | Fluorescent stain | Smart Label  |
|----------------------|-----------------|-------------------|--------------|
| Detection limit (ng) | 100 - 10        | 1 or less         | 1 or less    |
| Dynamic range        | E2              | E4-5              | E 4-5        |
| Staining/ destaining | yes             | yes               | not required |
| Reproducibility      | low             | low               | very high    |
| Protein monitoring   | no              | no                | yes          |

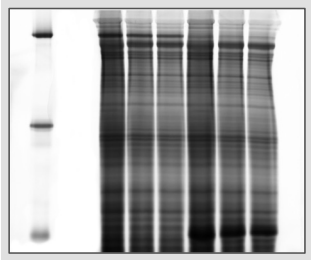


## The Smart Label

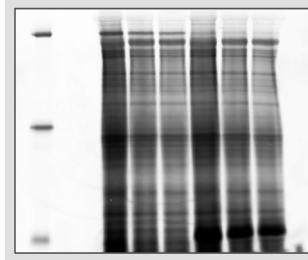
### Monitoring total protein during every step of the Western procedure

Protein transfer and subsequent Western blot procedure (e.g. washing steps with TBST) lead to inconsistent loss of total protein. The high sensitivity *Smart Label* precisely monitors the amount of protein for every protein band at every step of the Western blot procedure. Figures show sample protein (20  $\mu\text{g}$  per lane) visualized by using *Smart Labels* during Western blot analysis. Rapid protein detection was performed using an Octopus Fluorescence Imager.

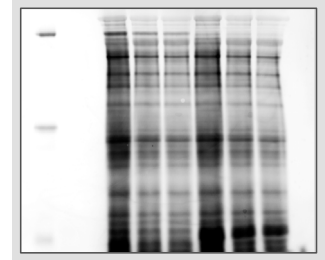
Gel



Blot immediately after transfer



Blot at moment of target detection

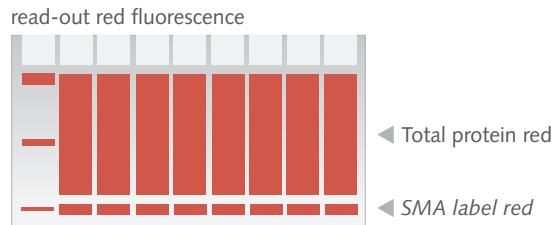
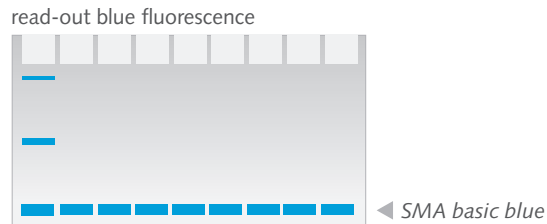


## The Smartalyzer

### Using a smart standard for accurate normalization

Providing standards to samples allows for precise normalization. The *Smartalyzer* (SMA) is a blue fluorescent standard which is added to every protein sample. The sample and the SMA are then labeled with reactive *Smart Label Red*.

After this procedure the sample protein fluoresces in red, SMA fluoresces in basic blue and in labeling red. SMA is available in two sizes: SMA S = 12.5 kDa, SMA L = 80 kDa.

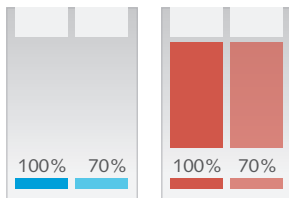


## The Smartalyzer

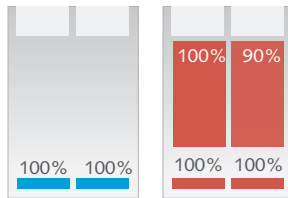
### Using a smart standard for accurate normalization

The relation between the blue and red fluorescent SMA and the corresponding red fluorescent sample protein allows for precise lane to lane protein normalization. This includes normalization of unequally loaded lanes due to loss of sample (figure A) or samples differing in their protein content (figure B). The labeling efficiency is monitored and if necessary normalized (figure C).

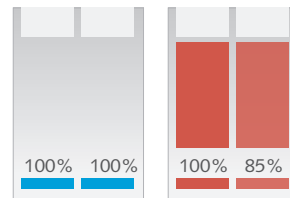
**A** Use SMA *basic blue* for loading normalization.



**B** Use protein label red for normalization of samples loaded by volume or incorrect protein determination.



**C** Use SMA *label red* for labeling efficiency normalization.



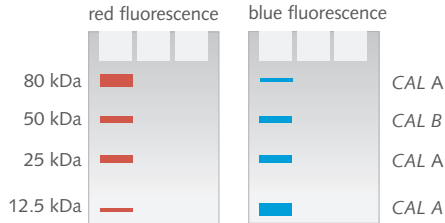
## The Calibrator

### The standard for experiment-to-experiment comparison

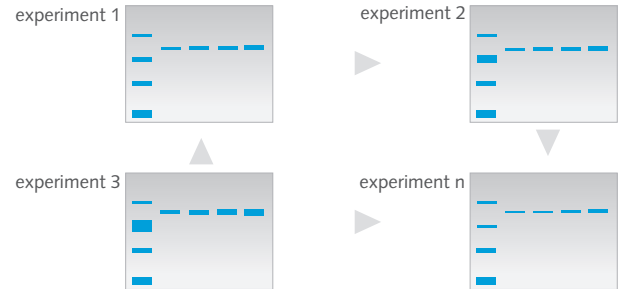
For comparison of different experiments, a bi-fluorescent standard, the *Calibrator* (CAL) is applied to the gel. CAL consists out of three markers (CAL A, each red and blue fluorescent) and one 50 kDa marker (CAL B) specific to the 2<sup>nd</sup> antibody used for these experiments.

In addition, CAL is a fluorescent protein weight marker.

Use CAL A as fluorescent marker

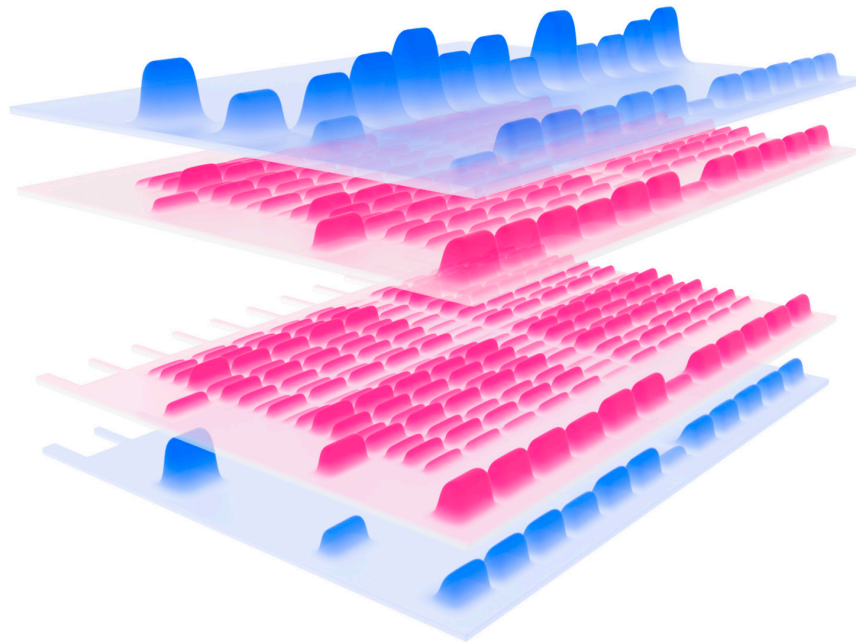


Use CAL A+B to compare different gels or blots



## Smart Protein Layers

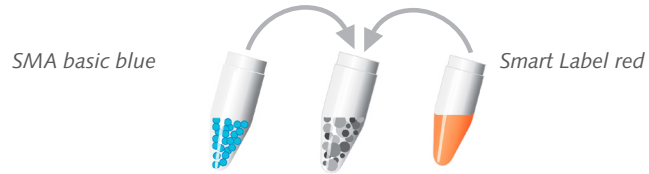
### Layer overview



# Smart Protein Layers Workflow

1

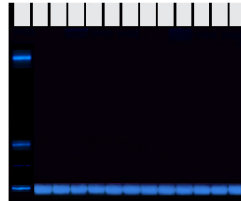
Add *Smart Label* & *Smartalyzer* (SMA) to your protein.



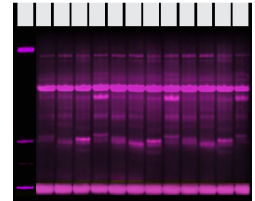
2

Run SDS-PAGE. Detect blue and red fluorescence.

- SMA basic blue
- CAL A



- Total protein
- SMA label red
- CAL A

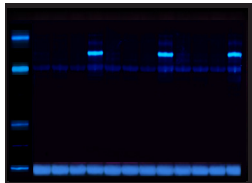


## Smart Protein Layers Workflow

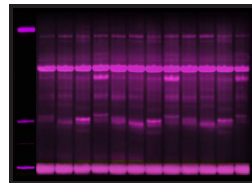
3

Perform Western blot and detect fluorescence of target (blue) and normalized total protein

- *SMA basic blue*
- *CAL A + B*



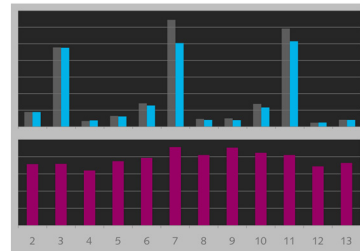
- Total protein
- *SMA label red*
- *CAL A*



4

Process data and evaluate.

- Detection and determination of band and lane raw volumes.
- Normalization of gel load (based on *SMA basic blue*).
- Normalization of *Smart Label* (based on *SMA label*).
- Normalization of target protein signal (based on total protein).
- Experiment-to-experiment normalization (based on *CAL*).

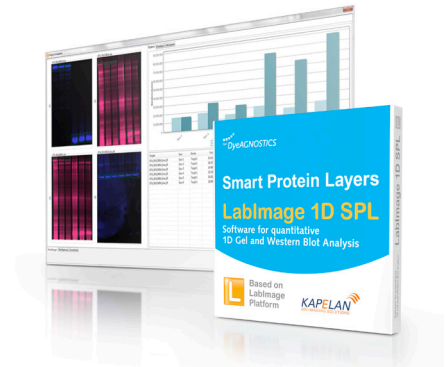


## SPL-LabImage Software

Fast and accurate data processing and evaluation

Accurate evaluation requires several data processing steps. The SPL-LabImage software is designed to determine band and lane raw volumes of an unlimited number of projects. It automatically uses *SMA basic color*, *SMA label color* and protein signal to perfectly normalize the total protein of gels and blots. Normalization of target protein expression can then be precisely performed.

- Workflow-based software
- Maximum flexibility (e.g. reference lane can be freely chosen)
- Runs on MAC or PC (English or German)





# Target detection

## Which SPL kit fits to my method of target protein detection?

The best method (in terms of signal stability, easy handling, exposure time) of your target protein detection depends on target abundance in the sample. Higher abundant proteins can be easily detected using first or secondary fluorescent antibodies. For most targets detection using Immuno blue is the ideal choice. However, very low abundant protein targets (femtogram range) can be detected only by using adequate femto chemiluminescence kits. The table below shows recommended combinations of SPL kits for total protein visualization and the optimal method for target protein detection.

| Target protein        | Detection method   | SPL kit  |
|-----------------------|--|----------|
| higher picogram range | fluorescent antibody conjugates<br>(preferably red or infra red) | SPL Blue |
| lower picogram range  | Immuno Blue fluorescence<br>+ HRP conjugated 2nd AB              | SPL Red  |
| femtogram range       | chemiluminescence kits<br>for femtogram range                    | SPL Red  |

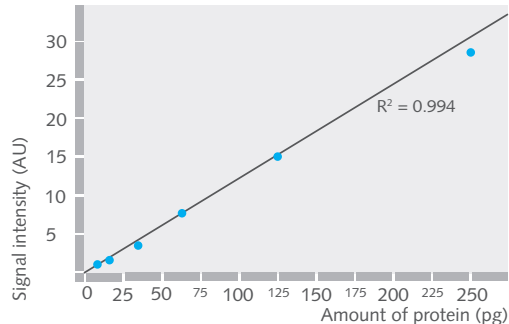
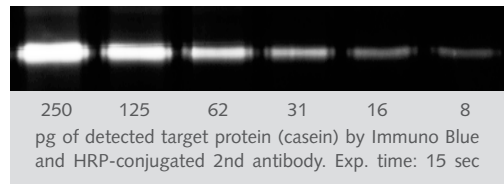
## Immuno Blue Fluorescent Substrate

### Target protein detection

Immuno Blue Fluorescent Substrate combines the high sensitivity of chemiluminescence with signal stability and the short exposure time of fluorescent antibody conjugates.

HRP-conjugated 2<sup>nd</sup> antibodies transform Immuno Blue into a fluorescent compound which precipitates and remains stable on the blot.

- Short exposure time (10 -100 x shorter than ECL)
- Long signal stability of several months
- Sensitivity in the lower pg range



## Fluorescence imaging requirements

Smart Protein Layers can be easily read-out with epi-blue (496/ 520 nm) and epi-red (650/ 665 nm) fluorescence imagers.

The following options are recommended:

- Red + blue fluorescence available:  
Choose SPL Red for total protein + Immuno Blue for target protein detection  
Choose SPL Blue for total protein + Smart Red 2<sup>nd</sup> AB for a highly abundant target
- Blue fluorescence + chemiluminescence available:  
Choose SPL Blue for total protein, chemiluminescence for target protein detection

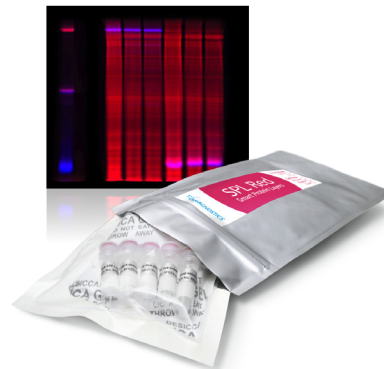
For further questions please contact [info@dyeagnostics.com](mailto:info@dyeagnostics.com)

## Product overview

### SPL Red for 40 Western blots

recommended in combination with Immuno Blue (PR 840)

- *Smart Label red*  
400 reactions for 1-20 µg of protein each
- *Smartalyzer basic blue*  
choose size S (12.5 kDa) or size L (80 kDa)
- *Calibrator red & blue*  
3 fluorescent marker bands, choose *Calibrator B* specific to 2nd antibody



| Kit description  | Smart Label | Smartalyzer      | Calibrator | Product No. |
|------------------|-------------|------------------|------------|-------------|
| SPL Red S Mouse  | red         | SMA basic blue S | Mouse      | PR911-M     |
| SPL Red S Rabbit | red         | SMA basic blue S | Rabbit     | PR911-R     |
| SPL Red S Goat   | red         | SMA basic blue S | Goat       | PR911-G     |
| SPL Red L Mouse  | red         | SMA basic blue L | Mouse      | PR912-M     |
| SPL Red L Rabbit | red         | SMA basic blue L | Rabbit     | PR912-R     |
| SPL Red L Goat   | red         | SMA basic blue L | Goat       | PR912-G     |

## Product overview

### SPL Blue for 40 Western blots

recommended in combination with Smart Red fluorescent 2<sup>nd</sup> antibody

- *Smart Label blue*  
400 reactions for 1-20 µg of protein each
- *Smartalyzer basic red*  
choose size S (12.5 kDa) or size L (80 kDa)
- *Calibrator red & blue*  
3 fluorescent marker bands, choose *Calibrator B* specific to 2nd antibody



| Kit description   | Smart Label | Smartalyzer     | Calibrator | Product No. |
|-------------------|-------------|-----------------|------------|-------------|
| SPL Blue S Mouse  | blue        | SMA basic red S | Mouse      | PR914-M     |
| SPL Blue S Rabbit | blue        | SMA basic red S | Rabbit     | PR914-R     |
| SPL Blue S Goat   | blue        | SMA basic red S | Goat       | PR914-G     |
| SPL Blue L Mouse  | blue        | SMA basic red L | Mouse      | PR915-M     |
| SPL Blue L Rabbit | blue        | SMA basic red L | Rabbit     | PR915-R     |
| SPL Blue L Goat   | blue        | SMA basic red L | Goat       | PR915-G     |

## Product overview

### Immuno Blue Fluorescent Substrate

for 40 Western blots 10 x 10 cm

Product No. PR 840

### Smart Red fluorescent 2<sup>nd</sup> Antibody

0.5 mg

Product No. PR 831 (for anti-mouse)

Product No. PR 832 (for anti-rabbit)

Product No. PR 832-G (for anti-goat)

### Blotting Kit

low fluorescent membrane + blotting paper,  
precut 10 x 10 cm, pack of 10

Product No. PR 811

### Beo Dry Blotter

high quality blotting of up to 18 mini gels, no buffers required

Product No. PR87

### SPL-LabImage Software

for quantitative 1D gel and Western blot analyses

Mac or PC, English or German version

Product No. PR 989

### Octoplus SPL Imager

red and blue fluorescence (optional: + green, + IR)

high end chemiluminescence detection







- + I want a stain-free protein visualization for gels and Western blots.
- + I want to precisely quantify and compare different blots.
- + And if my Western fails, I want to know why.

I am using Smart Protein Layers.

